

## **Evaluation of the Possible Effect of Methotrexate, Fluoxetine and Moringa Oleifera in Rat Model of Rheumatoid Arthritis**

Mohamed E. Mansour, Mohie ELDin Sherif, Yasmeen M. Ismail, Abeer Abd Elhameed, Sara A. Bassouiney

Department of clinical Pharmacology, Faculty of Medicine Benha University, Egypt.

Correspondence to: Sara A. Bassouiney, Department of clinical Pharmacology, Faculty of Medicine Benha University, Egypt.

#### Email:

saraawadbenha@gmail.com

Received: 3 January 2021

Accepted: 4 September 2021

### **Abstract**

**Background:** Rheumatoid arthritis (RA) is a chronic, relapsing inflammatory and autoimmune multisystem disease that affects the joints. Methotrexate (MTX) is considered by many rheumatologists to be the most important and useful disease-modifying antirheumatic drugs (DMARDs). Fluoxetine is a selective serotonin reuptake inhibitor that has been widely used for the treatment of depression due to its safe profile. Moringa Oleifera (MO) is known for its nutritional and medicinal uses. Various parts of these plant act as cardiac and circulatory stimulants. Aim of work: The present study was designed to evaluate the possible effect of methotrexate, Fluoxetine and Moringa Oleifera on rat model of RA. Materials and methods: Rats were classified into: Group I: control normal group. Group II: was not treated (diseased group). Group III: was treated with methotrexate (MTX) (0. 6 mg/kg/week/by oral gavage). Group IV: was treated with fluoxetine (20 mg/kg/day by oral gavage) Group V: was treated with Moringa Oleifera extract

(200 mg/kg/day by oral gavage). Treated groups received drugs for 4 weeks. **Results**: Treated groups showed significant improvement in all parameters (rheumatoid factor (RF), tumor necrosis factor alpha (TNF- $\alpha$ ), C-reactive protein (CRP), reduced glutathione (GSH), anti-cyclic citrullinated peptide (anti-CCP), arthritis score).and improvement of the histopathology of the joint. A significant reduction in the score was seen in the treated group at the end of 3<sup>rd</sup> and 4<sup>th</sup> weeks. With best results in Moringa Oleifera group in comparison with other treated groups. **Conclusion:** Methotrexate, fluoxetine and Moringa Oleifera extract groups showed significant improvement of parameters of RA.

**Key words**: Rheumatoid arthritis, methotrexate, fluoxetine, Moringa Oleifera, adjuvant-induced arthritis.

### **Introduction:**

Rheumatologic diseases are the most prevalent diseases worldwide; they are one of the main causes of disability and morbidity all over the world with greatly bad impact on the quality of life. (1).

They are characterized by the presence of long-standing inflammation of the joints resulting in symmetric polyarthritis and synovial membrane hypertrophy with progressive joint damage, bone and cartilage destruction and also deformity. The disease is systemic, leading to extraarticular manifestations (EAM) (2).

The pathogenesis of the rheumatoid joint involves Synovial hyperplasia results from a marked increase in macrophage and fibroblast-like synoviocytes. Locally expressed degradative enzymes, including metalloproteinase and proteases, digest the extracellular matrix and destroy the articular structures. (3).

Cytokines, particularly IL-1 and IL-17, tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and reactive oxygen intermediates affect chondrocytes that undergo apoptosis. This results in cartilage degradation and joint-space narrowing on radiography (4).

Systemic involvement may be explained by the systemic activities of cytokines released from the inflamed synovium. (5). The main classes of drugs are currently used: analgesics, non-steroidal antiinflammatory drugs (NSAIDs), glucocorticoids, as well as biologic and non-biologic disease-modifying antirheumatic drugs (DMARDs). (6).

Methotrexate (MTX) is the recommended first-line DMARD used in the treatment of RA. This is due to its low cost and many patients achieve disease control with monotherapy. However, about 30% of patients develop inadequate treatment response and many patients stop MTX due to toxicity.(7).

Part of MTX anti-inflammatory action is due to its folate antagonist Methotrexate up taken into the cells by folate transporter 1 (FOLT, also known as RFC1). Within the cell, MTX is polyglutamated in a reversible reaction by folylpolyglutamate synthase. Polyglutamation of methotrexate increasing inhibition of dihydrofolate reductase (DHFR), thymidylate synthetase and 5-aminoimidazole- 4-carboxamide ribonucleotide transformylase.(8).

The most common adverse effects include: gastrointestinal side effects (nausea, vomiting, abdominal pain) followed by stomatitis (oral ulcers), liver function abnormalities, bone marrow suppression, and alopecia. (9).

Neuropsychiatric manifestations are quite common in RA, including depression, cognitive dysfunction,

behavior changes, spinal cord compression and peripheral nerve involvement.(10).

The current trend of medical treatment of rheumatoid arthritis seeks for new drugs with more efficacies and less side effects since methotrexate, a standard diseased modified anti-rheumatoid drug, causes many adverse effects and toxicities. (11). Fluoxetine is a selective serotonin

reuptake inhibitor that has been widely used to enhance the neurotransmission of serotonin in the central nervous system and has emerged as the drug of choice for the treatment of depression due to is safer profile and fewer side effects. (12).

Also there are important functions of fluoxetine related to the central nervous system e.g. neuroprotection; anti-inflammatory properties, antioxidant properties and anti-apoptotic properties, with greater neuron survival and a reduction in apoptosis mediators and also oxidative substances, such as superoxide dismutase and hydrogen peroxide. (13).

Moringa Oleifera (MO) is known for its nutritional and numerous medicinal uses. Various parts of these plant act as cardiac and circulatory stimulants, possess antitumor, antipyretic, antiulcer, antispasmodic, diuretic, antihypertensive, analgesic, antioxidant, antimicrobial, and anti-inflammatory effect.(14).

### Aim of work:

These observations provided a rationale for testing the effect of monotherapy with fluoxetine and MO extract in comparison with MTX monotherapy on adjuvant induced arthritis (AIA).

### **Materials & methods:**

#### A. Animals:

The study is carried out on 30 Adult male albino rats obtained from Experimental Animal Breeding Farm, Helwan-Cairo) weighing between 150- 200 g (at the beginning of the study), were used for invivo experiments. They were acclimatized for one week and were caged (6 rat/ cage) fully ventilated room at room in the temperature pharmacology department, Benha Faculty of Medicine. Rats were fed a standard chow with water. This study was approved from ethical committee of Benha Faculty of Medicine.

### **B. Drugs and chemicals:**

Freund's adjuvant (CFA) Complete (Sigma-Aldrich Chemical Company), Methotrexate (Minapharm., Egypt), fluoxitine (Amoun pharmaceutical Co, Egypt)., Moringa olifera (powder) (National Research Centre, Giza, Egypt), Formalin, solution, neutral 10% formaline (El Gomhoria Pharmaceutical Chemical Co., ARE), Urethane, Ethyl carbamat, white crystals (Sigma Chemical Co., USA), Hematoxylin and eosin (E.Merk, Darmastadt.,)[Germany], Rheumatoid factor kits (Abnova corporation, Taipei city ,Taiwan).,CRP kits (Thermoscientific, USA), Tumor necrosis factor alpha (TNF-α) kits (USA & Canada | R&D Systems, Inc), Reduced Glutathione kits (Biodiagnostic Co., Giza, Egypt), anticyclic citrullinated peptide (anti-CCP)kits (Alpha diagnostic international,Texas,USA)

#### C. Induction of RA:

Complete Freund's Adjuvant Arthritis was induced by S.C injection of 0.4 ml of compelete Freund's adjuvant in the right hind limb for 12 day in three doses (one dose every four days).(11).

### D. Experimental design:

### **Group** (1): **Non-arthritic** untreated normal control group:

This group received a standard chow and tap water with no medication.

# Group (2): Untreated rats with complete freund's adjuvant arthritis group:

This group was injected with complete freund's adjuvant to induce rheumatoid arthritis.

### Group (3): Rheumatoid arthritis (RA) methotrexate treated group:

This group received a standard chow and tap water with methotrexate at a dose (0. 6 mg/kg/week/by oral gavage) (15) for 4 weeks after induction of arthritis with CFA.

### Group (4): Rheumatoid arthritis (RA) rats, fluoxetine treated group:

This group treated with fluoxetine (20 mg/kg/day in saline by oral gavage) (16) for four weeks after after induction of arthritis with CFA.

### Group (5): Rheumatoid arthritis (RA) rats, Moringa oleifera treated group:

This group treated with (200 mg/kg/day by oral gavage of methanolic extract of M. oleifera) (17) for four weeks after induction of arthritis with CFA.

The treated groups received drugs for 4 weeks. Dose selection was based on previously published studies and pilot experiments.

At the end of study period, blood samples were collected from the retro-orbital venous plexus of rats using microcapillary tubes. (18).

The blood samples (each=2ml) were allowed to clot at room temperature, centrifuged at 3000 rotation/minute and the sera were separated. Samples were stored at -20  $\acute{C}$  in dark containers for measurement of rheumatoid factor (RF), C-reactive protein (CRP), tumor necrosis factor alpha (TNF- $\alpha$ ), reduced glutathione (GSH)and anti-cyclic citrullinated peptide (anti-CCP).

Rats were euthanized at the end of the study and hind paws were removed. They were embedded in paraffin after fixing in formalin solution (10% neutral buffered).

Sections were cut in various slices having thickness of 6 um and examined under microscope after staining with for hematoxylin-eosin perivascular inflammatory cell infiltrate in synovium, morphological changes including synovial cell hyperplasia and proliferation, villous hyperplasia, inflammatory cells infilterations and dilated blood vessels.(19).

#### E. Assessment of arthritis:

a- Rats were scored for arthritis (arthritis index) by a set visual criterion at the end of each week of experiment according to the following criteria: No change = 0, Erythema = 1, Mild swelling = 2, Gross swelling = 3, Gross swelling and deformity = 4. (20).

**b-** Microscopic examination of sections of paw joint stained by H&E stain.

### F. Biochemical assays:

- a) RF was determined by ELISA technique (21).
- b) Serum TNF-α was measured by ELISA (22).
- c) Serum CRP was determined using an enzyme-linked immunosorbent assay
   (23).
- d) Serum GSH was determined by using a colorimetric method (24).
- e) Serum anti-cyclic citrullinated peptide (anti-CCP) was determined using an enzyme-linked immunosorbent assay.(25).

### **Statistical analysis:**

The clinical data were recorded on a report form. These data were tabulated and analysed using the computer program SPSS (Statistical package for social science) version 26 to obtain descriptive statistics were calculated for the data in the form of mean and standard deviation for quantitative data.

In the statistical comparison between the different groups, the significance of difference was tested using ANOVA test Used to compare mean of more than two groups of quantitative data.

A P value <0.05 was considered statistically significant (\*) while >0.05 statistically insignificant P value <0.01 was considered highly significant (\*\*) in all analyses.

### **Results:**

Injection of (CFA) resulted in significant increase in level of RF, CRP, TNF- $\alpha$  and anti-CCP antibodies compared to control normal group.

There is a significant progressive increase in arthritic score of diseased group every week without treatment compared to the score at 1st day before adjuvant injection. While there was significant decrease in GSH level compared to control normal group. Joints obtained from diseased group showed proliferated blood vessels,

inflammatory infiltrate and pannus formation.

Regarding, the treated groups there was significant improvement in serum RF, TNF- $\alpha$ , CRP, GSH and anti-CCP antibodies compared to diseased group. MTX showed better results than fluoxetine

group, also MO showed the best results in all treated groups in the following parameters serum RF, CRP and anti-CCP antibodies. While fluoxetine showed more decrease in TNF- $\alpha$  and increase in GSH more than MTX group but the best improvement is also in MO group.

**Table (1):** The effect of treatment with MTX, fluoxetine and MO on (serum RF, TNF-α, CRP, GSH and anti-CCP antibodies) on experimentally induced rheumatoid arthritis, in rats:

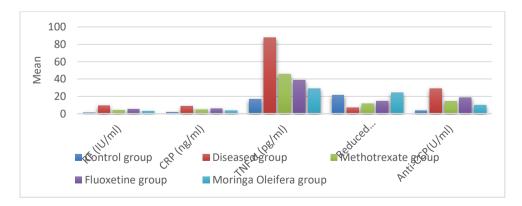
	C41		D:I		Methot	rexate	Fluox	etine	Mor	inga
	Control group	Diseased group		group		group		Oleifera group		
	Mean	±SD	Mean	±SD	Mean	±SD	Mean	±SD	Mean	+SD
RF	1.46	0.16	0.62-	0.70	3.94	0.20	5.14	0.22	2.91	0.10
(IU/ml)	1.46	0.16	9.62 <b>a</b>	0.70	ab	0.38	abc	0.23	abcd	0.19
CRP	1.64	0.11	0.625	0.54	4.74	0.76	5.9	0.22	3.67	0.20
(ng/ml)	1.64	0.11	8.63 <b>a</b>	0.54	ab	0.76	abc	0.32	abcd	0.38
TNF-α	16.67	0.50	97.520	0.54	45.84	0.70	38.64	0.00	29.16	0.64
(pg/ml)	16.67	0.50	87.53 <b>a</b>	0.54	ab	0.78	abc	0.98	abcd	0.64
Reduced					11 62		140		24.15	
Glutathione	21.47	0.87	6.97 <b>a</b>	0.34	11.63	0.66	14.8	0.76	24.15	1.04
(μg/ml)					ab		abc		abcd	
Anti-CCP	3.71	1.19	29.05 <b>a</b>	0.35	14.3	1.57	18.85	0.54	10.12	1.04
(U/ml)					ab		abc		abcd	

a: significant versus G1 at p<0.05

b: significant versus G2 at p<0.05

c: significant versus G3 at p<0.05

d: significant versus G4 at p<0.05



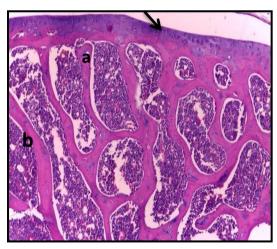
**Figure (1):** The effect of treatment with MTX fluoxetine and MO, on (serum RF, TNF- $\alpha$ , CRP, GSH and anti-CCP antibodies) on experimentally induced rheumatoid arthritis, in rat

Table (2): The effect of treatment with fluoxetine and methotrexate, on arthritic score at different times,	on
experimentally induced rheumatoid arthritis in rats:	

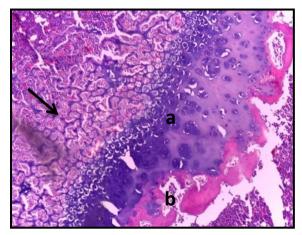
	G1		G2		G3		G4		G5	
	Mean	±SD	Mean	±SD	Mean	±SD	Mean	±SD	Mean	±SD
Arteritis score 0	0.0	0.0	3.0	0.0	3.0	0.0	3.0	0.0	3.0	0.0
Arteritis score 1 <sup>st</sup> w	0.0	0.0	4.0 <b>a</b>	0.0	3.33 <b>ab</b>	0.52	3.5 <b>ab</b>	0.55	3.0 <b>ab</b>	0.0
Arteritis score $2^{nd}$ w	0.0	0.0	4.0 <b>a</b>	0.0	3.0 <b>ab</b>	0.63	3.33 <b>ab</b>	0.52	2.5 <b>abd</b>	0.55
Arteritis score 3 <sup>rd</sup> w	0.0	0.0	4.0 <b>a</b>	0.0	2.83 <b>ab</b>	0.75	3.0 <b>abc</b>	0.63	2.33 <b>abcd</b>	0.52
Arteritis score 4 <sup>th</sup> w	0.0	0.0	4.0 <b>a</b>	0.0	2.10 <b>ab</b>	0.63	2.67 <b>abc</b>	0.52	1.5 <b>abcd</b>	0.55

a: significant versus G1at p<0.05 b: significant versus G2 at p<0.05 c: significant versus G3 at p<0.05 d: significant versus G4 at p<0.05

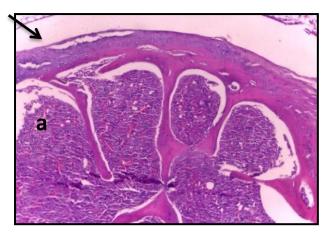
### **Histopathological changes:**



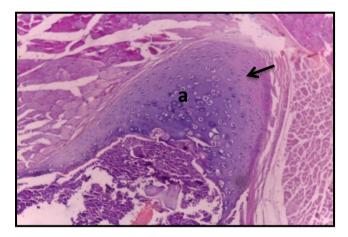
**Figure (2):** Photomicrograph of a cut section in normal rat joint shows (black arrow) flattened synovial membrane (a) cartilage associated with (b) bone marrow with no inflammatory cellular infiltrate (H&E x 100).



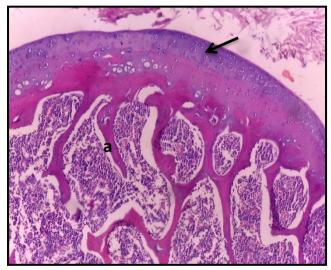
**Figure (3):** Photomicrograph of a cut section in rat joint of diseased group shows inflammatory infiltrate and proliferated blood vesseles (black arrow), synovial hyperplasia(a), pannus formation(b) (H&E x 100).



**Figure (4):** Photomicrograph of a cut section in rat joint of MTX treated group shows improvement of the inflammatory response (a) and flattening of synovial membrane(black arrow). (H&E x100).



**Figure (5):** Photomicrograph of a cut section in rat joint of fluoxetine treated group showed (black arrow) multilayered synovial membrane, (a) mild improvement of inflammatory infiltrate (H&E x100).



**Figure (6):** Photomicrograph of a cut section in rat joint of Moringa oleifera treated group shows (black arrow) marked flattening of superficial synovial layer (a) and marked improvement of inflammatory infiltrate (H&E x 100).

Regarding arthritis score, a significant reduction in the score was seen in treated groups at the end of 3rd and 4th weeks compared to diseased group, MO group was better than fluoxetine and MTX groups.

Histopathology revealed improvement of the joint as regard synovial hyperplasia, cartilage degeneration and inflammatory cell infiltration. This improvement was observed in all treated groups with best result in MO group

### **Discussion:**

The present work was designed to evaluate the effect of Methotrexate, Fluoxetine and Moringa Oleifera extract for 4 weeks, on rheumatoid factor (RF, tumor necrosis factor-  $\alpha$  (TNF- $\alpha$ ), C-reactive protein (CRP), reduced glutathione (GSH), anticyclic citrullinated peptide (anti-CCP) and arthritis score and histopathological of joint, changes on RA induced experimentally in rats by S.C. injection of complete Freund's adjuvant in right hind limb.

Complete Freund's adjuvant (CFA)-induced arthritis is considered a scientifically standard experimental procedure for the induction of chronic immune-pathological RA in laboratory animals with similar cellular immunity

response and pathological mechanism as in the human. (26).

The present study showed that S.C. injection of (CFA) resulted in increase in level of RF, CRP, TNF- $\alpha$ , arthritic score, decrease in level of GSH. This finding is similar to the observations of other study proved that injection of CFA led to significant elevation of serum RF, serum TNF- $\alpha$  and decrease blood GSH level compared to normal control group (27).

These results are in agreement with the study that reported that injection of complete Freund's adjuvant leads to edematous inflammation, increased vascularity owing to vasodilation, marked inflammatory cell infiltration compared to normal control group (28).

CFA showed that serum rheumatoid factor and serum CRP level were significantly increased in CFA rats compared with normal control group which is in consistence with our study (29).

The results in this study also are parallel with the study that tested the effect of galantamine on adjuvant-induced arthritis in rats. The adjuvant arthritis model showed the anti-CCP level increased in the untreated adjuvant arthritic rats relative to the healthy control group. (30).

The cytokines are involved in the pathogenesis of RA. In particular, tumor necrosis factor alpha (TNF $\alpha$ ) has been

suggested as one of the most potent cytokines associated with RA. (31).

TNF-α induces activation of leukocyte and endothelial and synoviocyte activation and survival, cytokine and chemokine amplification, angiogenesis and nociceptor activation. The blockade of TNF significantly decreases the production of other pro-inflammatory cytokines and chemokines, as IL-1, IL-6, IL-8, or GMCSF (32).

Reduced glutathione (GSH) is the most abundant intracellular small-molecule thiol and is essential for maintaining the thiol status of various molecules. GSH has many biological roles, including protection against reactive oxygen and nitrogen species (ROS/NOS), which are reduced by two GSH molecules forming oxidized glutathione (GSSG) in the process (33).

Methotrexate significantly improved all the tested parameters of rheumatoid arthritis, arthritic score and histopathology of the joints. The result of this study is in agreement with the observations of other study that reported that MTX treatment after induction of RA by CFA, induced a highly significant decrease in serum RF, serum TNF-α, serum CRP and induced a highly significant increase in blood GSH level compared with diseased group.(34). Also, in line with these results another study reported that the paw edema, RF and

anti-CCP antibody were significantly reduced with MTX treated group (35).

In addition, our results showed that a significant reduction in paw diameters in MTX-treated was that observed in arthritic group and histopathology of joints of MTX-treated group showed reduction in vascular proliferation, destruction of cartilage, synovial membrane, and sub periosteal region (36). For the arthritic score another study who aimed to compare the safety effectiveness of MTX administrated his study is in agreement with our results as administration decreased the MTX arthritis score significantly (37)

The anti-inflammatory actions of MTX are also due to the participation of adenosine. Adenosine is an endogenous antiinflammatory factor in arthritis. MTX acts 5-aminoimidazole-4-carboxamide ribonucleotide suppressor and increases adenosine levels. Adenosine suppresses neutrophil migration areas inflammation, promotes the differentiation of macrophages and also inhibits the production of interleukin-1 or leukotriene B4 (38).

Some researchers have postulated that MTX treatment decrease of the severity of arthritis by down-regulation of proinflammatory TNF-α, IL-6 and IL-17A cytokine expression (39).

Regarding treatment with fluoxetine, it showed significant improvement of RF, CRP, **GSH** TNF-α. and anti-CCP antibodies, compared to diseased. For arthritic score, this group showed improvement of the score at the end of 3rd and 4th weeks, compared to diseased group.

The effect of fluoxetine on serum level of TNF- $\alpha$  and arthritis score was studied and showed significant improve in arthritis score and significant decrease in TNF- $\alpha$  level compared to non-treated group as approved in our study.(40).

Fluoxetine inhibiting effect on cytokine secretion can be explained by the study that reported that fluoxetine counteracting depressive symptoms by inhibiting the reuptake of serotonin and thus, augments serotonin concentration. The relatively high extracellular serotonin levels can inhibit the secretion of cytokines (41).

Also a possible mechanism by which fluoxetine inhibit endosomal Toll like receptor (TLR) such as TLR 8 which plays an important role in the production of TNF- $\alpha$ .( 40).

Fluoxetine effect on serum CRP was significantly decreased as shown by the study of the effect of fluoxetine on inflammatory markers; it showed significant reduction in serum CRP level, and this run in consistence with the current work (42).

IL-6 is also the main pro-inflammatory cytokine that induce synthesis of type 1 acute phase proteins such as CRP, elevated levels of stress leads to activation that triggers an NF-KB-dependent cascade of pro-inflammatory events that contribute to increases in CRP (43).

It also proved that fluoxetine decrease expression of nuclear factor NF- $\kappa B$ , this explain the reduction of the release of a number of pro-inflammatory and cytotoxic factors such as TNF- $\alpha$ , IL-1 $\beta$ , nitric oxide, and reactive oxygen radicals. It also suggested that fluoxetine inhibits the mRNA for these cytokines (as well as for IL-6) (44).

Our study is in line with the study that showed fluoxetine has increased level of GSH significantly as approved in our study (45).

Effect of fluoxetine on GSH is explained by restoring the affected GSH pathways with Fluoxetine treatment may relate to neuroprotection, as the antioxidative effects of fluoxetine are thought to be mediated by increases in serotonin levels (46).

And it has been reported that fluoxetine suppressing T cell proliferation and inhibit interferon- (IFN) production in whole blood cultures. These also explain the decreased cytokines level in our present study (47).

In addition fluoxetine may also inhibit the response of antigen presenting cells these can explain the decreased levels of auto-antibodies as RF and Anti-CCP antibodies (48).

According to paw edema and arthritis score fluoxetine showed significant reduction in edema and improvement of arthritis score especially at 3rd and 4th weeks and this is in line with y the anti-inflammatory and immunomodulatory effects of fluoxetine in rat models.(49).

Histopathological examination to fluoxetine group showed decreased inflammatory infiltration, bone and cartilage destruction which is in line with study that approved the inhibitory effect of fluoxetine on inflammation and bone loss in rats. And this is due to the antiinflammatory effect of fluoxetine (50 & 51).

On the other hand another study showed that 10 mg/kg of fluoxetine the same dose in our study showed a small reduction in the clinical score and a slower decrease in paw swelling but at the higher dose (25 mg/kg), fluoxetine profoundly halted disease progression, with no further elevation in the clinical score or paw swelling (40).

Regarding treatment with Moringa oleifera extract, it showed highly significant improvement of RF, TNF-α, CRP, GSH and anti-CCP, compared to diseased,

normal control group and other treated groups. For arthritic score, this group showed significant improvement of the score at the end of 3rd and 4th weeks, compared to diseased group and other treated groups.

The result of this study showed that MO group has highly significant decrease in TNF-  $\alpha$  and this is supported by a study which studied anti-inflammatory effect of MO extract, it showed MO reduce TNF- $\alpha$  significantly and this may explained by  $\beta$ -sitosterol present in MO which is a compound with potent activity against inflammation, whose mechanism of action includes reducing the production of TNF- $\alpha$  (52).

MO also used in treatment of asthma and associated allergic diseases; these studies showed significant decrease in TNF- $\alpha$ , IL-6, these findings also indicate that the possible mechanism of action may be associated with a reduction in cytokine production/release (53).

Lipopolysaccharide (LPS) can bind to TLR and activate the NF- $\kappa$ B signaling pathway. The activation of these cascades and transcription factors subsequently results in the releasing of proinflammatory cytokines by macrophages and circulating monocytes, resulting in a transient immune activation, which is characterized by elevated levels of TNF- $\alpha$ , IL-1 $\beta$ , and IL-6, MO extract strongly

inhibit the LPS-induced expression of IL-6 and TNF- $\alpha$  during inflammation.(54).

In addition, our results are in consistence with the study that showed the effect of MO on vascular oxidative stress in hypertensive rats, it showed that significant decrease in CRP in MO treated group. (55).

IL-6 is an important mediator of the inflammatory response as it participates in the development and differentiation of B-and T-cells, as well as the activation of acute phase proteins as CRP, MO extract inhibit mRNA expression of IL-6and thus consciously leads to decreased CRP level.(56).

Also the study of the antioxidant, antiinflammatory and anti-arthritic effect of M. oleifera showed in histopathology inflammation. minimal no pannus formation and erosion of epithelial cells. Also decrease in paw edema of Moringa treated rats as approved in our study. (57) M.O extract shows significant protection against lymphocytic infiltration, bone destruction and cartilage erosion and this is in line with our study. Also significant reduction in RF and TNF-α, and this is supported by the study of the protective effect of ethanolic extract of seeds of MO in arthritic rats (58).

B cells isolated from RA synovium can secrete RF and anti-CCP antibodies.

indicating that the autoantibody is produced locally in the joint (59).

Moringa seed extract has the ability to attenuate the chronic immune-mediated inflammatory responses typical of certain diseases such as asthma and RA and this explain the decrease in parameters of rheumatoid arthritis such as RF and anti-CCP antibodies (60).

MO extract showed significant increase in GSH and this is supported by the study of the protective effect of MO extract seeds against diabetic nephropathy in rats (61).

The significant increase in GSH is also supported by studying the effect of MO seed extract on induced testicular toxicity in rats (62).

These significant increase in GSH may explained by Glutathione Reductase (GR) is essential in maintaining adequate GSH level by facilitating the regeneration of GSH from oxidized glutathione (GSSG). The protective effect of MO was also reflected in the induction of GR activity by MO extract. (63).

Cysteine and methionine rich proteins that are present in high amounts in MO seeds .Beside this, MO, which is also rich with other potent antioxidant s like vitamin C, vitamin E and B-carotene.(64)

### **Conclusion**

Methotrexate, Fluoxetine and Moringa Oleifera had improved adjuvant arthritis. Moringa Oleifera extract can be used as new treatment in cases of rheumatoid arthritis thus can decrease MTX dose to avoid its side effects. Also, fluoxetine can be used in in treatment of rheumatoid arthritis to avoid possible side effect of high doses of methotrexate also it has beneficial effect in cases of depression associated with rheumatoid arthritis.

### **Sources of funding**

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

### References

- Goma, S. H., Mahran, D. G., El-Hakeim, E. H., Ghandour, A. M., Abdelaziz, M. M., Galal, M. A., et al., (2016). Spectrum of Rheumatic Diseases in Egypt is Similar/Different from that in Non-Arabic Countries: An Inpatient Comparison, volume 1 | Issue 1.
- 2) Amaya J, Botello-Corzo D, Calixto J. (2012): Usefulness of patients-reported outcomes in rheumatoid arthritis focus group. Arthritis. 2012:935(187).
- 3) **Firestein, G. S. (2003).** Evolving concepts of rheumatoid arthritis. Nature, 423(6937), 356-361.
- **4) McInnes, I. B., & Schett, G. (2011).** The pathogenesis of rheumatoid arthritis. New England Journal of Medicine, 365(23), 2205-2219.
- Nishimoto, N., & Kishimoto, T. (2006). Interleukin 6: from bench to bedside. Nature clinical practice Rheumatology, 2(11), 619-626.
- 6) Kumar P and Bain k
  (2013):Pharmacotherapy Options in
  Rheumatoid Arthritis ,Clinical Medicine
  Insights Arthritis Musculoskelet Disorders
  35–43.
- 7) Ling, S. F., & Bluett, J. (2020). Pharmacogenetics of methotrexate response

- in rheumatoid arthritis: an update 3-6, volume 21 , Issue 1.
- 8) Brown, P. M., Pratt, A. G., & Isaacs, J. D. (2016). Mechanism of action of methotrexate in rheumatoid arthritis, and the search for biomarkers. Nature Reviews Rheumatology, 12(12), 731-742.
- 9) Singh, R. G., Negi, P. S., & Radha, C. (2013). Phenolic composition, antioxidant and antimicrobial activities of free and bound phenolic extracts of Moringa oleifera seed flour. Journal of functional foods, 5(4), 1883-1891.
- **10) Joaquim, A. F., & Appenzeller, S. (2015).** Neuropsychiatric manifestations in rheumatoid arthritis. *Autoimmunity reviews*, *14*(12), 1116-1122.
- 11) Hendawy, O. M.; Ahmed, W. M. S.; Abosaif, A. A. (2015): Effect of atorvastatin and vitamin D on freund adjuvant-induced rheumatoid arthritis in rat. J Bioequiv Availab., 7:090-094. doi: 10. 4172/jbb. 100221.
- 12) Ravera, S., Ramaekers, J. G., & de Gier, J. J. (2012). Are selective serotonin reuptake inhibitors safe for drivers? What is the evidence?. Clinical therapeutics, 34(5), 1070-1083.
- 13) Caiaffo V. Oliveira B., de Sa´ F. & Neto J. (2016): Anti-inflammatory, antiapoptotic, and antioxidant activity of fluoxetine. Pharmacology Research Perspective, 4(3).
- **14)** Ravindra A., Priya S. and Siddheshwar S.(2019): A pharmacological Review On Moringa Oleifera , World Journal of Pharmaceutical Research ,8,910-920.
- 15) Rovenskýa, J., Švíka, K., Stancíkováa, M., Ištoka, R., Ebringerb, L., & Ferencíkc, M. (2006): The effects of selenium enriched enterococus faecium M-74 on methotrexate treatment of rats with adjuvant arthritis. International Journal of Probiotics and Prebiotics, 1(2):137-144.
- 16) Branco-de-Almeida, L. S., Franco, G. C., Castro, M. L., dos Santos, J. G., Anbinder, A. L., Cortelli, S. C., et al., (2012). Fluoxetine inhibits inflammatory response and bone loss in a rat model of ligature-induced periodontitis. *Journal of periodontology*, 83(5), 664-671.
- 17) <u>Mahajan</u> S., <u>Mali</u> R. & <u>Mehta</u> A.(2008):Protective Effect of Ethanolic

- Extract of Seeds of *Moringa oleifera* Lam. Against Inflammation Associated with Development of Arthritis in Rats. <u>Journal of Immunotoxicology</u>. Pages 39-47, volume 4,issue 1.
- **18) Schemer, S. (1967):** The blood morphology of laboratory animals.3rd ed. Philadelphia,USA: Davis company;42.
- 19) Faisal, R., Ahmad, N., Fahed, Y. S., & Chiragh, S. (2018). Anti-arthritic effect of thymoquinone in comparison with methotrexate on pristane induced arthritis in female Sprague Dawley Rats. *Journal of Ayub Medical College Abbottabad*, 30(1), 3-7
- 20) Vetal, S., Bodhankar, S. L., Mohan, V., & Thakurdesai, P. A. (2013): Anti-inflammatory and antiarthritic activity of type –A procyanidine polyphenols from bark of Cinnamomumzeylanicum in rats. Food Sci Hum Wellness, 2:5967.
- 21) Engvall, E. and Perlman, p. (1971): Enzyme-Linked Immunosorbent Assay, (ELISA) Quantitative Assay of Immunoglobulin G. Immunochemistry. 8: 871-874.
- 22) Kasumagic-Halilovic, E.; Prohic, A. and Cavaljuga, S. (2011): Tumor necrosis factoralpha in patients with alopecia areata. Indian Journal of Dermatology, 56(5):494-6.
- 23) Nathan, B. R. and Scheld, W. M. (2002): The potential roles of C-reactive protein and procalcitonin concentrations in the serum and cerebrospinal fluid in the diagnosis of bacterial meningitis. Curr Clin Top Infect Dis., 22:155-165.
- 24) Beutler, E.; Duron, O.; Kelly, M. B. (1963): Glutathione reduced. J Lab Clin Med.; 61 .882-888.
- 25) Avčin, T., Cimaz, R., Falcini, F., Zulian, F., Martini, G., Simonini, G., et al.,(2002). Prevalence and clinical significance of anticyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. Annals of the rheumatic diseases, 61(7), 608-611.
- 26) Mahdi, H. J., Khan, N. A. K., Asmawi, M. Z. B., Mahmud, R., Vikneswaran, A., &Murugaiyah, L. (2018). In vivo antiarthritic and anti-nociceptive effects of ethanol extract of Moringaoleifera leaves on complete Freund's adjuvant (CFA)-induced

- arthritis in rats. Integrative Medicine Research, 7(1), 85-94.
- 27) Refaat, R., Salama, M., Meguid, E. A., El Sarha, A., & Gowayed, M. (2013): Evaluation of the effect of losartan and methotrexate combined therapy in adjuvant-induced arthritis in rats. Eur. J. Pharmacol., 698: 421-428.
- 28) Saleem, A., Saleem, M., Akhtar, M. F., Shahzad, M., &Jahan, S. (2020). Moringarivae leaf extracts attenuate complete Freund's adjuvant-induced arthritis in Wistar rats via modulation of inflammatory and oxidative stress biomarkers. Inflammopharmacology, 28(1), 139-151.
- 29) Mehta, A., Sethiya, N. K., Mehta, C., & Shah, G. B. (2012). Anti–arthritis activity of roots of Hemidesmusindicus R. Br. (Anantmul) in rats. Asian Pacific Journal of Tropical Medicine, 5(2), 130-135.
- 30) Gowayed, M. A., Refaat, R., Ahmed, W. M., & El-Abhar, H. S. (2015). Effect of galantamine on adjuvant-induced arthritis in rats. European journal of pharmacology, vol.,764, 547-553.
- 31) Kojima, A., Kobayashi, T., Ito, S., Murasawa, A., Nakazono, K., &Yoshie, H. (2016). Tumor necrosis factor-alpha gene promoter methylation in Japanese adults with chronic periodontitis and rheumatoid arthritis. Journal of periodontal research, 51(3), 350-358.
- **32) Noack, M., &Miossec, P. (2017).**Selected cytokine pathways in rheumatoid arthritis.In Seminars in immunopathology (Vol. 39, No. 4, pp. 365-383).Springer Berlin Heidelberg.
- 33) Damgaard, D., Bjørn, M. E., Steffensen, M. A., Pruijn, G. J., & Nielsen, C. H. (2016).Reduced glutathione as a physiological co-activator in the activation of peptidylargininedeiminase.Arthritis research & therapy, 18(1), 102.
- 34) Makar, N., Elhawary, A. H., Emam, H., Abo Ria, N., &Shaaban, E. E. S. (2020). Possible Beneficial Effect of Metformin Alone or in Combination with Methotrexate in Rheumatoid Arthritis Induced Rat Model. *Benha Medical Journal*, *37*(1), 143-154.
- 35) Ali, Z., Atia, H., El Allawy, R., &AbdAlla, S. (2017). Anti-hepatotoxic and synergistic effects of sesame oil with methotrexate in adjuvant induced arthritis. *The Egyptian*

- Journal of Biochemistry and Molecular Biology, 35(1-2), 77-92.
- 36) Roy, T., Banerjee, I., Ghosh, S., Dhali, R. S., De Pati, A., &Tripathi, S. K. (2017). Effects of co-treatment with pioglitazone and methotrexate on experimentally induced rheumatoid arthritis in Wistar albino rats. *Indian journal of pharmacology*, 49(2), 168.
- 37) Wang, X., Yan, X., Wang, F., Ge, F., & Li, Z. (2018): Role of methotrexate chronotherapy in collagen-induced rheumatoid arthritis in rats. Z Rheumatol., 77(3):249-255.
- **38)** Koyama, A., Tanaka, A., & To, H. (2017). Daily oral administration of low-dose methotrexate has greater antirheumatic effects in collagen-induced arthritis rats. Journal of Pharmacy and Pharmacology, 69(9), 1145-1154.
- 39) Zhao, P. W., Jiang, W. G., Wang, L., Jiang, Z. Y., Shan, Y. X., & Jiang, Y. F. (2014). Plasma levels of IL-37 and correlation with TNF-α, IL-17A, and disease activity during DMARD treatment of rheumatoid arthritis. PloS one, 9(5), e95346.
- 40) Sacre, S., Medghalchi, M., Gregory, B., Brennan, F., & Williams, R. (2010). Fluoxetine and citalopram exhibit potent antiinflammatory activity in human and murine models of rheumatoid arthritis and inhibit toll-like receptors. Arthritis & Rheumatism, 62(3), 683-693.
- 41) Lu, Y., Ho, C. S., Liu, X., Chua, A. N., Wang, W., McIntyre, R. S., et al., (2017). Chronic administration of fluoxetine and proinflammatory cytokine change in a rat model of depression. *PloS one*, 12(10), e0186700.
- **42)** Coccaro, E. F., Lee, R., Breen, E. C., & Irwin, M. R. (2015).Inflammatory markers and chronic exposure to fluoxetine, divalproex, and placebo in intermittent explosive disorder.Psychiatryresearch, 229(3), 844-849.
- 43) Chavda, N., & ND Kantharia, J. (2011). Effects of fluoxetine and escitalopram on Creactive protein in patients of depression. Journalofpharmacology&pharmacotherapeuti cs, 2(1), 11.
- 44) Liu, D., Wang, Z., Liu, S., Wang, F., Zhao, S., & Hao, A. (2011). Anti-inflammatory effects of fluoxetine in lipopolysaccharide

- (LPS)-stimulated microglial cells. Neuropharmacology, 61(4), 592-599.
- **45) Perić, I., Stanisavljević, A., Gass, P.,** & Filipović, **D.** (2017). Fluoxetine reverses behavior changes in socially isolated rats: role of the hippocampal GSH-dependent defense system and proinflammatory cytokines. European Archives of Psychiatry and Clinical Neuroscience, 267(8), 737-749.
- **46) Zafir, A., Ara, A., &Banu, N.** (2009). In vivo antioxidant status: a putative target of antidepressant action. Progress in Neuro-Psychopharmacology and Biological Psychiatry, 33(2), 220-228.
- **47) Diamond, M., Kelly, J. P., & Connor, T. J.** (**2006**). Antidepressants suppress production of the Th1 cytokine interferon-γ, independent of monoamine transporter blockade. European Neuropsychopharmacology, 16(7), 481-490.
- **48)** O'neill, L. A. (2008). Primer: Toll-like receptor signaling pathways—what do rheumatologists need to know. Nature clinical practice Rheumatology, 4(6), 319-327.
- 49) Kostadinov, I., Delev, D., Petrova, A., Stanimirova, I., Draganova, K., Kruzliak, P. et al., (2015). Study on anti-inflammatory and immunomodulatory effects of fluoxetine in rat models of inflammation. *European Journal of Inflammation*, 13(3), 173-182.
- **50)** Roumestan, C., Michel, A., Bichon, F., Portet, K., Detoc, M., Henriquet, et al., (2007). Anti-inflammatory properties of desipramine and fluoxetine. *Respiratory research*, 8(1), 35.
- **51) Abdel-Salam, O. M., Baiuomy, A. R., & Arbid, M. S. (2004).** Studies on the anti-inflammatory effect of fluoxetine in the rat. Pharmacological research, 49(2), 119-131.
- 52) Araújo, L. C. C., Aguiar, J. S., Napoleão, T. H., Mota, F. V. B., Barros, A. L. S., Moura, M. C., et al., (2013). Evaluation of cytotoxic and anti-inflammatory activities of extracts and lectins from Moringaoleifera seeds. *PloS one*, 8(12), e81973.
- 53) Mahajan, S. G., Banerjee, A., Chauhan, B. F., Padh, H., Nivsarkar, M., & Mehta, A. A. (2009). Inhibitory effect of n-butanol fraction of Moringaoleifera Lam. seeds on ovalbumin-induced airway inflammation in a guinea pig model of asthma. International journal of toxicology, 28(6), 519-527,

- 54) Luetragoon, T., PanklaSranujit, R., Noysang, C., Thongsri, Y., Potup, P., Suphrom, N., et al., (2020). Bioactive compounds in Moringaoleifera Lam. leaves inhibit the pro-inflammatory mediators in lipopolysaccharide-induced human monocyte-derived macrophages. Molecules, 25(1), 191.
- 55) Randriamboavonjy, J. I., Rio, M., Pacaud, P., Loirand, G., & Tesse, A. (2017). Moringaoleifera seeds attenuate vascular oxidative and nitrosative stresses in spontaneously hypertensive rats. *Oxidative Medicine and Cellular Longevity*, 2017, 17(41), 294.
- 56) Abdel-Daim, M. M., Khalil, S. R., Awad, A., Abu Zeid, E. H., El-Aziz, R. A., & El-Serehy, H. A. (2020). Ethanolic Extract of Moringaoleifera Leaves Influences NF-κB Signaling Pathway to Restore Kidney Tissue from Cobalt-Mediated Oxidative Injury and Inflammation in Rats. Nutrients, 12(4), 1031.
- 57) Saleem, A., Saleem, M., & Akhtar, M. F. (2020). Antioxidant, anti-inflammatory and antiarthritic potential of Moringa oleifera Lam: An ethnomedicinal plant of Moringaceae family. South African Journal of Botany, 128, 246-256.
- 58) Mahajan, S. G., Mali, R. G., & Mehta, A. A. (2007). Protective effect of ethanolic extract of seeds of Moringaoleifera Lam. against inflammation associated with development of arthritis in rats. *Journal of Immunotoxicology*, 4(1), 39-47.
- 59) Reparon-Schuijt, C. C., van Esch, W. J., van Kooten, C., Schellekens, G. A., de Jong, B. A., van Venrooij, W. J., et al.,

- (2001). Secretion of anti–citrulline containing peptide antibody by B lymphocytes in rheumatoid arthritis. Arthritis & Rheumatism: Official Journal of the American College of Rheumatology, 44(1), 41-47.
- 60) Leone, A., Spada, A., Battezzati, A., Schiraldi, A., Aristil, J., &Bertoli, S. (2016). Moringaoleifera seeds and oil: characteristics and uses for human health. International Journal of Molecular Sciences, 17(12), 2141.
- 61) Abdalrhman, I, K., Naziha, N. &Ali, M. (2018) Evaluation the Protective Effect of Moringa Oleifera Seeds Oil Against Diabetic Nephropathy in Male Rats. Scientific Journal of Specific Education and Applied Sciences, (1)1,267-282.
- 62) Abarikwu, S. O., Benjamin, S., Ebah, S. G., Obilor, G., &Agbam, G. (2017). Oral administration of Moringaoleifera oil but not coconut oil prevents mercury-induced testicular toxicity in rats. Andrologia, 49(1), e12597.
- 63) Uma, N., Jr, Fakurazi, S., &Hairuszah, I. (2010). Moringaoleifera Enhances Liver Antioxidant Status via Elevation of Antioxidant Enzymes Activity and Counteracts Paracetamol-induced Hepatotoxicity.Malaysian journal of nutrition, 16(2), 293–307.
- **64) Gupta, R., Dubey, D. K., Kannan, G. M.,** & **Flora, S. J. S.** (2007).Concomitant administration of Moringaoleifera seed powder in the remediation of arsenic-induced oxidative stress in mouse. Cell Biology International, 31(1), 44-56.

**To cite this article:** Mohamed E. Mansour, Mohie ELDin Sherif, Yasmeen M. Ismail, Abeer Abd Elhameed, Sara A. Bassouiney. Evaluation of the Possible Effect of Methotrexate, Fluoxetine and Moringa Oleifera in Rat Model of Rheumatoid Arthritis. BMFJ 2022:39 (academic issue):35-51.

Benha medical journal, vol. 39, Special issue (academic), 2022