

Evaluation of MicroRNA-210 (miR-210) as a Diagnostic and Prognostic Biomarker in Pre-eclampsia Pregnancies

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Abstract:

Objective: This study aims to assess the level of miR-210 in Egyptian women with pre-eclampsia (PE) and to evaluate its role in diagnosis and prognosis of the disease. **Subjects and Methods:** The study was conducted on 30 pregnant women with PE divided into two groups: group (I): included 15 cases with mild PE and group (II): included 15 cases of severe PE and, 20 healthy pregnant women with matched age and sex were included as control group. All women included in the study were subjected to, history taking full clinical examination, laboratory investigations included (CBC), (PT), (ALT, AST), (urea, creatinine), detection of protein in urine, as well as miRNA-210 gene expression by RT-PCR. **Results:** Patients with PE showed a highly significantly increase in serum miR-210 (P value <0.001) compared to control as well as, it was higher in severe PE than in mild PE (P value <0.001). MiR 210 have highly significant positive correlation with (systolic, diastolic, MABP), (Proteinuria) and (PT) a significant positive correlation with (AST), (ALT),.However no significant correlation was found with (Hb%) ,platelet count, PTT and INR, serum urea and creatinine. PTT was statistically significant (P value 0.002).The best cutoff value of PE (2.03) with sensitivity and specificity of 90.0% and 85.0% respectively. **Conclusion:** Expression of mir-210 is upregulated in pre-eclampsia and was higher in severe than in mild. Hence, the serum miR-210 can be used as a diagnostic, prognostic biomarker in PE patients and understanding pathophysiology.

Key Words: Pre-eclampsia , miR-210 , RT_PCR

Introduction

Pre-eclampsia is a multi-system disorder that is unique to human pregnancy, affecting 2-5% of pregnancies worldwide and leads to maternal deaths in 10-15% (1).

Pre-eclampsia can manifest as either a maternal syndrome (hypertension and proteinuria with / without other multi-system dysfunction) or fetal syndrome.(2).

Although the etiology of pre-eclampsia is thought to be related to insufficient spiral arterial conversion leading to decreased uteroplacental perfusion and, therefore, is thought to give rise to chronic placental ischemia and hypoxia (3).

MicroRNAs are members of a larger class of non-coding RNAs that control gene expression and regulate biological processes by targeting mRNA and inducing translational repression or RNA degradation. MicroRNAs are on average a~70nt-long transcript (4). MicroRNAs expression affects several human cellular function from proliferation and differentiation to apoptosis and associating with miRNAs pathological alteration in diseases from cancer to myocardial infarction (5). miR-210 is associated with a variety of important processes like cell cycle regulation, cell

survival, differentiation, angiogenesis, as in metabolism as in hypoxic conditions and cancer (6),(7).

Over expression of miR-210 can regulate cell response to hypoxia through mitochondrial function promoting the shift from mitochondria respiration to glycolysis and induce angiogenesis (8). Also in placental tissue isolated from pre-eclamptic patients shows miR-210 expression is increased (9).

Previous studies have demonstrated that the dysregulation of the specific plasma-related miRNAs expression is associated with the development of PE (1) and (10).

Aim of the study

This study aimed to assess the level of miR-210 expression in the serum of Egyptian women with pre-eclampsia and to evaluate its role in diagnosis and prognosis of PE.

Subjects

This case/control study was conducted on 30 pregnant women with PE divided into two groups: group (I): included 15 cases with mild PE, group (II): included 15 cases of severe PE .In addition, 20 healthy pregnant women with matched age and sex representing the control group (III).

PE patients were admitted to Gynecology and Obstetrics Department in Benha University Hospitals. The study was conducted from the period of (January 2018 to January 2019.) after approval of the local ethical committee of Benha University. Informed written consent was obtained from each participant.

PE patients was classified According to a study done in 2018:, mild PE was diagnosed when blood pressure $\geq 140/90$ mmHg on 2 occasions, at least 6 hours apart, but without evidence of end-organ damage, in a woman who was normotensive before 20 weeks' gestation, and the systolic blood pressure (SBP) increased by 30 mmHg or diastolic blood pressure (DBP) increased by 15 mmHg. Positive proteinuria after 20 weeks of gestation.

Severe PE was diagnosed when SBP of ≥ 160 mmHg or DBP of ≥ 110 mmHg on 2 occasions at least 6 hours apart in a normotensive pregnant woman before pregnancy. Proteinuria of more than (5 g) in a 24-hour collection And Presence of symptoms of severe PE were detected such as pulmonary edema, cyanosis, oliguria and/or impaired liver function, thrombocytopenia, oligohydramnios,

decreased fetal growth, or placental abruption.(11).

Exclusion criteria:

- Fetal congenital malformations or chromosomal abnormalities.
- Recent infection.
- Anti-phospholipid syndrome.
- Trauma.
- Drug or alcohol abuse.
- Preexisting hypertension.
- Thrombophilia with history of PE.
- Receiving anticoagulant.

Methods

All cases included in the study were subjected to:

- 1- Full history taking: such as personal and maternal history including: age, gravidity, positive family history, gestational age and onset of preeclampsia.
- 2- Clinical examination: General and clinical examination including: BP, Mean Arterial Pressure (MABP), edema, weight, height, and body mass index (BMI).

3- Laboratory investigation:

Blood sample:

- 6 mL of venous blood was drawn under complete aseptic conditions:

- CBC by 1 mL blood was mixed with EDTA used, by automated cell counter (Sysmex KX 21N; Sysmex, Inc., Mundelein, IL, USA).
- Coagulation profile (PT, PTT, INR) by 1.6 ml was added on (Na citrated tube for) using automated coagulometer (Coatron A4, Teco, Germany).
- The rest of blood was withdrawn into plain tube and separated serum was aliquoted and stored at -80°C until used. The stored serum then thawed and used for:
 1. Biochemical assays: liver function tests and kidney function tests using Biosystem A15 auto-analyzer (Biosystems S.A., Barcelona, Barcelona, Spain).
 2. miRNA-210 detection by RT-PCR.

Urine sample: 24 h Urine sample was collected & centrifuged for protein estimation by a turbidimetric method (using TCA 3%, by Biosystem reagent kit using

semi-automated analyzer BTS-350 (Biosystem S.A., Barcelona, Spain).)

- Methods of miRNA-210 expression level by RT- PCR.

1-Purification of total RNA, including miRNA(Extraction).

QIAzolLysis Reagent was used to extract total RNA including microRNA from samples using miRNeasy Mini Kit (cat. no. 217004) (Qiagen,Germany) according to the manufacturer's protocol. QIAzolLysis Reagent was added to serum samples to facilitate lysis. After addition of chloroform, the lysate was separated in to aqueous and organic phases by centrifugation. RNA partitions were at the upper aqueous phase, RNA was extracted, and ethanol was added to provide suitable binding conditions for all RNA molecules. High-quality RNA was eluted in a small volume of RNase-free water.

Detection of RNA purity: by Nanodrop 2000 spectrophotometer (USA).

The ratio of absorbance at 260 nm and 280 nm is used to assess the purity of RNA. A ratio of nearly 2.0 is generally accepted as pure for RNA. If it is lower, this indicates protein, phenol or other contaminants that absorb strongly at or near 280 nm. It is

affected by change in sample acidity, wavelength accuracy of spectrophotometer and nucleotide mix in the sample.

260/230 ratio: Secondary measure of nucleic acid purity. Expected values are in the range of 2.0-2.2 (12).

2- Reverse transcription for quantitative real-time PCR (1st step):

cDNA was produced using miScriptII RT Kit (Cat.no.217004 QIAGEN, Germany) according to manufacturer instructions. The reverse transcription master mix was prepared using miScript HiSpec Buffer, miScript Nucleics Mix and miScript Reverse Transcriptase Mix. MiScript HiSpec Buffer converts mature miRNAs and certain small nucleolar RNAs and small nuclear RNAs selectively in to cDNA. The volume of RNA was different in each sample due to different concentrations so to use equal amounts of total RNA, the samples were diluted in RNase-free water. Template RNA and RNase-free water (12µL) was added to each tube containing reverse transcription master mix (8µL) forming final volume 20µL for each reaction in 0.2 mL polymerase chain reaction (PCR) tubes. Then mixed gently and briefly centrifuged. Tubes were then incubated at 37 °C for 60 min. followed by

incubation at 95 °C for 5 mins. In Veriti thermal cycler Applied Biosystem (USA).

3-Real-time PCR for detection of miRNA-210 (2nd step):

Quantification of miRNA was performed using miScript SYBR Green PCR Kit supplied by (QIAGEN, Germany) (cat.no 217004) according to the manufacturer's protocol. Each reaction was performed in a final volume of 25µl holding 2.5µl of cDNA, 2.5µl of miScript universal primer, 2.5µl of miScript primer assay (RNU 6 or miRNA 210), 5µl of RNase free water and 12.5 µl SYBR Green PCR Master mix. Forward and reverse miRNA specific primers were supplied by (Qiagen, Germany). MiR-210-5P primers, 5'-AGCCCCUGCCCACCGCACACUG -3' and RNU6, 5'- CTCGCTTCGGCAGCACA (F) -3'

Real-time PCR was performed on Real-time PCR cycler Applied Biosystems (USA) under the following conditions: Initial activation step at 95 °C for 15 min, followed by 40 cycles of denaturation at 94°C for 30s., annealing at 55°C for 1 min. and extension at 70°C for 1 min., in which fluorescence was acquired.

Normalization controls: the normalization control used was RNU6-2.

Analysis:

Detection and quantification of each gene was expressed as relative miRNA level compared with a standard housekeeping gene (RNU6-2) Selection of reference gene was done after evaluating its stability using the comparative delta-CT method (13).

The assessment of the obtained data from the miRNA expression was done by using the cycle threshold (Ct) method. The cycle threshold (Ct) is defined as the number of cycles required for the fluorescent signal to cross the threshold in RT-PCR. The expression level of the miRNA was reported as Δ Ct value, the fold change in the expression level of the miRNA was calculated (fold change = $2^{-\Delta\Delta Ct}$) (13).

Statistical Methods

Data management and statistical analysis were done using *SPSS vs.25*. (IBM, Armonk, New York, United states).

Comparisons between groups were done using either *Mann Whitney U* test; for numerical data, or *Chi-square* test; for categorical data. Mir210 level was compared between different degrees of severity and

controls using *Kruskal Wallis test*. *Post hoc* analysis was done which were all *Bonferroni* adjusted.

Correlation analyses were done between Mir210 and other parameters using Spearman's correlation. “r”,. Which ranges from -1 to +1. -1 indicates strong negative correlation, while +1 indicates strong positive correlation, and 0 indicates no correlation.

ROC analysis was done for Mir210 in diagnosing preeclampsia. (AUC), the best cutoff point, diagnostic indices including sensitivity, specificity, PPV and NPV were calculated. All P values were two sided. P values less than 0.05 were considered significant.

Results

The general characteristics of the studied groups was shown in table (1) shows no significant difference between (PE patients) and (control groups) regarding their maternal age, gestational age, body mass index and parity state (p value>0.05). Systolic, diastolic, mean arterial blood pressure), (AST),(Proteinuria) and (Mir-210)were highly significant increased in (PE patients) and (ALT)was significantly

increased than controls with P value (<0.05) .

Table (1):Comparison of demographic and laboratory data between the studied groups.

		Cases(n = 30)	Controls (n = 20)	P value
Maternal age (Years)	Mean ±SD	29 ±4	28 ±3	0.305
Gestational age (Wks)	Mean ±SD	30 ±5	33 ±3	0.243
Body Mass Index	Mean ±SD	33.96 ±4.22	31.66 ±6.28	0.096
Parity state	PG n (%)	23 (76.7)	16 (80.0)	0.780
	MG n (%)	7 (23.3)	4 (20.0)	
SBP (mmHg)	Mean ±SD	163 ±22	112 ±10	<0.001
DBP (mmHg)	Mean ±SD	108 ±11	76 ±6	<0.001
MABP (mmHg)	Mean ±SD	126 ±14	87 ±7	<0.001
Hemoglobin (gm/dl)	Mean ±SD	10.1 ±1.5	10.3 ±1.3	0.677
Platelets(1000/ul)	Mean ±SD	199 ±55	206 ±37	0.422
PT (Sec)	Mean ±SD	13.94 ±2.82	13.54 ±1.47	0.812
INR	Mean ±SD	1.19 ±0.23	1.12 ±0.12	0.26
PTT (Sec)	Mean ±SD	31 ±4.3	35.9 ±5.2	0.002
Urea (mg/dl)	Mean ±SD	42 ±15	39 ±16	0.307
Creatinine (mg/dl)	Mean ±SD	1.02 ±0.33	0.93 ±0.31	0.364
AST (U/L)	Median (range)	37 (11 - 300)	24 (12 - 112)	0.003
ALT (U/L)	Median (range)	52 (11 - 230)	36 (27 - 139)	0.025
Proteinuria (mg/24h)	Median (range)	1802 (341-10863)	89 (43 - 143)	<0.001
Mir210 -5p (fold)	Median (range)	6.01(1.62 - 18.62)	1.17(1 - 6.91)	<0.001

Mann Whitney U test was used for numerical data. *Chi-square test* was used for categorical data. (P <0.05) was statistically significant and (P <0.005) as highly significant.

The expression level of Mir210 was highly significant increase in PE patients compared to control group (P<0.001). MiR-210 was significantly higher in patients with PE than controls (P value : < 0.001).It significantly

higher in severe PE compared to patients with mild PE (P value : < 0.001) as shown in table (2) . Mir-210 showed a statistically highly significant correlation (positive correlation with systolic, diastolic and mean

arterial blood pressure, PT and Proteinuria) according to table (3) . ROC analysis was done for using Mir210 to diagnose preeclampsia. It revealed a significant Area Under Curve (AUC) of 0.933 with 95% confidence interval ranged from 0.863 to

1.0. The best cutoff value for diagnosis of PE was 2.03 with sensitivity and specificity of 90.0% and 85.0% respectively and PPV & NPV of 90.0% and 85.0% respectively. P value was (<0.001) as in figure (2).

Table (2): Mir210 level in all studied groups:

	Severe		Mild		Controls		P value
	Mean	±SD	Mean	±SD	Mean	±SD	
Mir210 (fold)	11.52	3.68	3.47	1.17	1.83	1.51	<0.001

Kruskal Wallis test was used. *Post hoc* analysis was done and different letters indicate significant pair . All post hoc were *Bonferroni* adjusted.

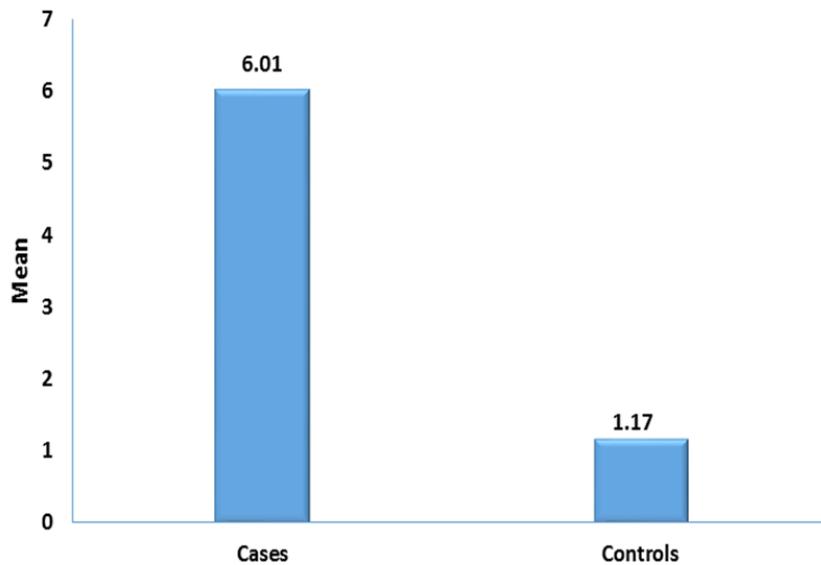


Figure (1): Mir-210 in both groups

Table (3): Correlation coefficient (r) between Mir210 and the study parameters

Mir210	r	P value
Maternal age (Years)	0.012	0.951
Body Mass Index	0.006	0.975
Gestational age (Weeks)	-0.203	0.282
Systolic blood pressure (mmHg)	0.402*	0.027
Diastolic blood pressure (mmHg)	0.473**	0.008
Mean Arterial Pressure	0.505**	0.004
Hemoglobin (gm/dl)	0.269	0.151
Platelets(1000/ul)	-0.017	0.928
PT (Sec)	0.493**	0.006
PTT (Sec)	-0.162	0.394
INR	0.262	0.162
Urea (mg/dl)	-0.1612	0.396
Creatinine (mg/dl)	-0.097	0.6122
AST (U/L)	0.214	0.256
ALT (U/L)	0.187	0.324
Proteinuria(mg/24h)	0.755**	<0.001

*Spearman's correlation was used, r = Correlation coefficient, *Significant, **highly significant.*

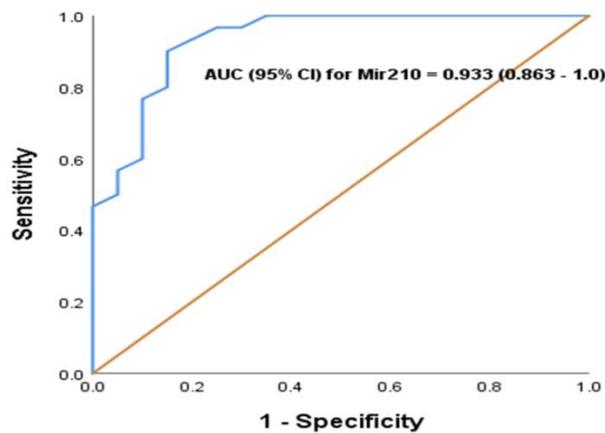


Figure (2): Roc curve of Mir210 in diagnosing pre-eclampsia

Discussion

miR-210 was identified in placenta of pregnant women with PE that has been suggested to induce repression of trophoblast cell invasion (14).

As for laboratory data of the groups, there was no significant difference between patients and controls regarding hemoglobin level or platelet count (P value = 0.677 & 0.422, respectively). The same was for PT and INR (P value = 0.812 & 0.26, respectively), however, PTT was statistically significantly shorter in patients compared to controls, (P value = 0.002) and it was within normal range.

PTT of PE patients compared with control group in the current study was not in agreement with other studies who observed that values of PTT were higher in the severe pre-eclamptic patients compared to controls (15) and (16). In another study it was found that sPE patients had abnormal coagulation profile (17), it was explained by a certain degree of coagulative dysfunction which occurs in the endogenous coagulative pathways of sPE, while exogenous coagulative pathways do not change (18).

There was no significant difference between patients and controls regarding serum urea

and creatinine (P value = 0.307, 0.364 respectively). But, ALT and proteinuria levels were highly significantly increased and AST was significantly elevated in patients than controls (P value = 0.003, <0.001, 0.025 respectively).

Other studies are agreed with our study noticed that significant increase in serum value of serum bilirubin and liver enzymes between preeclampsia cases compared to healthy pregnant, which explained by the effect of hypoxia of liver cells leading to necrosis of hepatocytes. (19) and (20), that increase was during the first 20 weeks of pregnancy which associated with higher risk for the development of severe preeclampsia in the second half of the pregnancy (21), that increased probability of maternal and fetal complications (22).

Regarding proteinuria, which was highly significantly elevated with PE in our study (P value <0.001), in agreement with other studies, this elevation is due to reversible kidney damage (23 and 24).

As regards miR-210, we found higher expression level in preeclampsia patients compared to controls with a high significant value (P value <0.001), which agreed with

other studies that confirming the role of elevated serum miR-210 as non-invasive method in diagnosis, prognosis and identifying women at-risk for monitoring and treatment through contribute to trophoblast function. (25) and (26). Other studies clarified that the expression levels of serum miR-210 was elevated starting from the second trimester in pregnancies complicated with PE.(1),(27)and(28)

Also other studies were similarly to our study reported that expression of placental miR-210 was up-regulated in patients with pre-eclampsia, due to inhibit the migration and invasion ability of trophoblast cells by placental miR-210,which regulated by transcriptional factor HIF-1 as well as NF- κ Bp50 leading to decreased STAT6and IL-4 .(29), (30) and (31).

The miR-210 expression may contribute to the occurrence of PE by interfering with potassium channel modulatory factor 1 -mediated signaling in the human placenta. (32), which they studied possible regulatory mechanisms by system biology approaches Through identifying regulated genes related to PE; targeted by human miR-210 .(33)

It was explained that high miR-210 expression is a possible modulator of mitochondrial dysfunction during PE. (34);

and also observed that hypoxia markedly increased miR-210 expression in PE placenta. (35). However, miR-210 was among the32 up-regulated key microRNAs and genes in PE (36).

In the present study, miR-210 was highly significantly elevated in patients with severe PE compared to both patients with mild PE and to control group (P value >0.001).These results suggested an association between miR-210 expression level and the severity of PE. In agreement of this study (37)

miR-210 in this study showed a significant positive correlation with SBP, DBP and MABP, PT as well as with proteinuria (P Value = 0.027, 0.008, 0.004, 0.006, and <0.001 respectively). In the same point of view, the urine miR-210 had a positive correlation with 24-hour urine proteins confirming the possibility of using urine miR-210 in evaluating the severity of kidney damage among PE patients. (1)

In addition, non-significant negative correlation was found in current study between mir210 and gestational age, platelet count, PTT, urea and creatinine level. (P value= 0.282, 0.928, 0.394, 0.396 & 0.6122 respectively). Moreover, non-significant positive correlation was found between mir210 and maternal age, BMI, hemoglobin

level, INR, AST& ALT level (P value = 0.931, 0.975, 0.151, 0.162, 0.256 & 0.324, respectively). ROC analysis was done for using mir210 for diagnosing preeclampsia. It revealed a significant AUC of 0.933 with 95% confidence, interval ranged from 0.863 to 1.0. miR-210 had sensitivity and specificity of 90.0% and 85.0% respectively with PPV and NPV of 90.0% and 85.0% respectively, which in agreement with the results of ROC curve were (AUC was 0.750 with 95% of confidence) (1).

Conclusion

Expression of mir-210 is upregulated in pregnant women with pre-eclampsia and it was found higher in severe than in mild preeclampsia. So, the serum miR-210 can be used as a non-invasive diagnostic and prognostic biomarker in pregnant women with pre-eclampsia. More studies with large sample size still needed to be investigated by increasing the sample size and its role in pathophysiology of the PE.

Abbreviations:

APTT = activated partial thromboplastin time,

AUC = area under the curve,

PE = pre-eclampsia,

PT = partial thromboplastin time,

miRNA = microRNA,

HIF-1 =hypoxic inducible factor.

NF-κB =nuclear factor kappa B

qRT-PCR = quantitative reverse transcriptase-polymerase chain reaction,

ROC = receiver operating characteristic.

* The authors declare that they have no conflict of interest.

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